



Effect of wall shear rate on wall flux of bacteria

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Industrial
partners



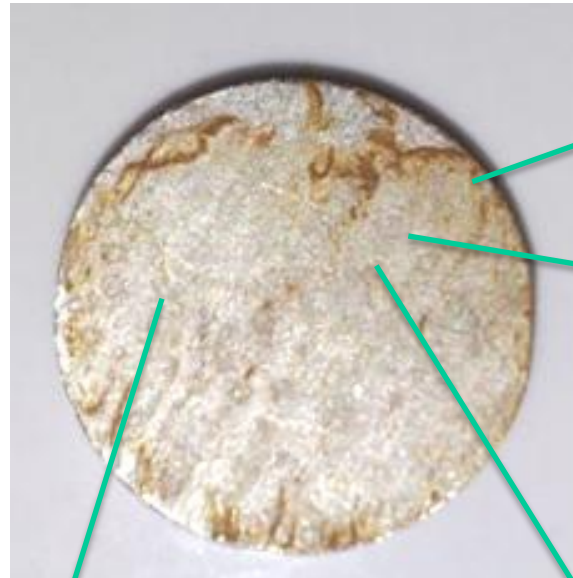
COST FP1005 "Fibre Suspension Flow Modelling" - ERCOFTAC SIG43 "Fibre suspension flows".
Coimbra – March 2013

Introduction

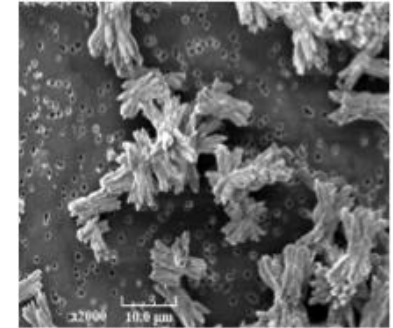
- Bacteria in drinking water : migration to the wall -> biofim ?
- Formation mechanisms of biofilm ? How to investigate?
- Test sections
- Transport equations
- Results
- Bacteria adhesion at the wall (cohesion of biofilm)
- Cleaning the surface colonized by biofilm
- Conclusions

Bacteria and deposits = Biofilm

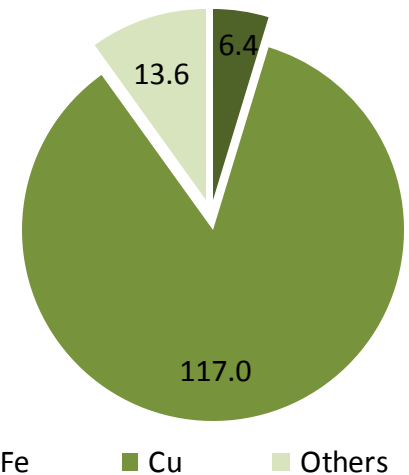
Drinking water bacteria and deposits



Carbonates



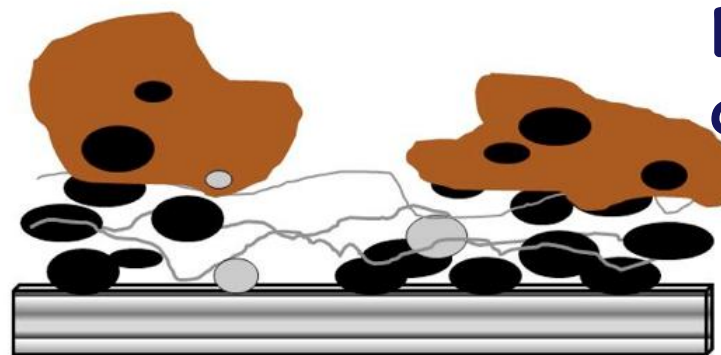
Metals ($\mu\text{g}/\text{cm}^2$)



Multispecies : Bacteria, fungi, ...

Brown deposits

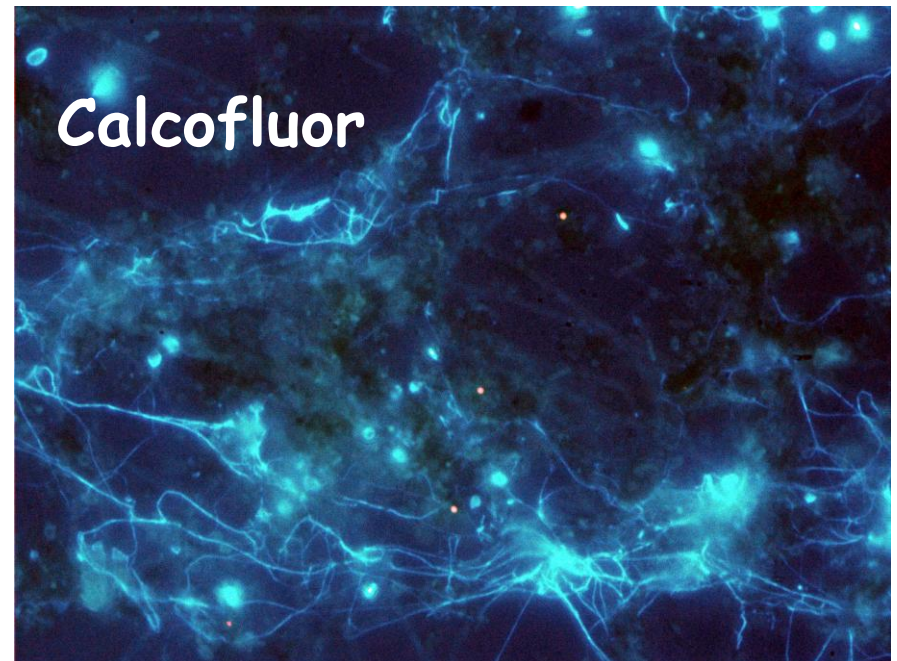
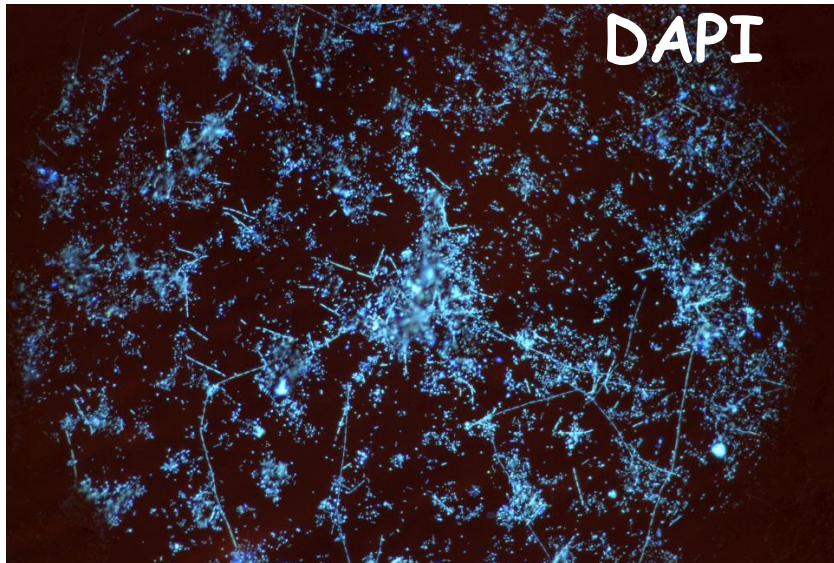
Bacteria



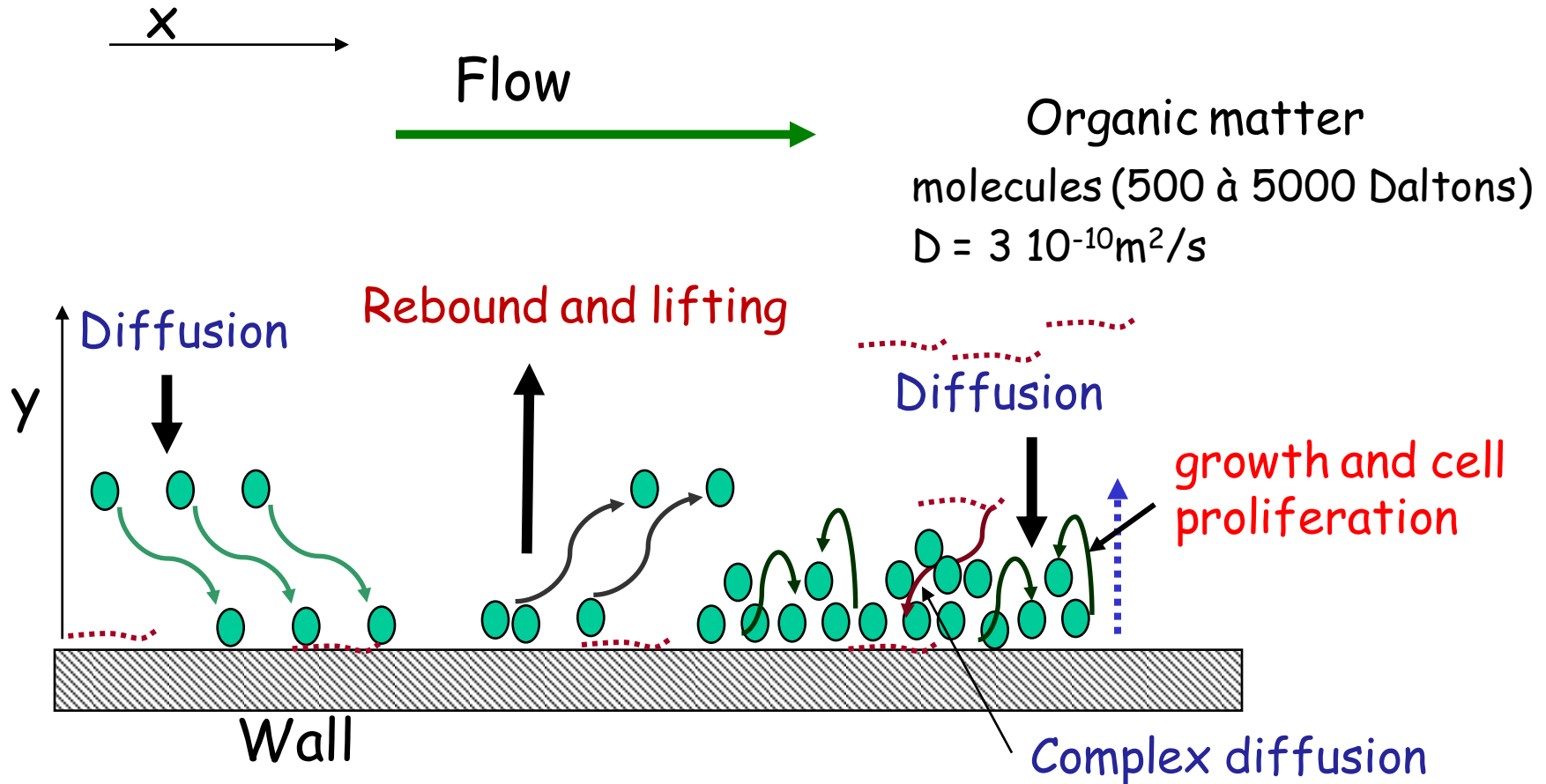
Organic matter :

20 to 100 $\mu\text{g TOC}/\text{cm}^2$

Drinking water biofilms (4 months old)



Wall activities



Phenomenological variables - mass transfer

Bacteria

$$D \approx 310^{-13} \text{ m}^2 / \text{s}$$

Close the wall

$$Pe = \frac{\gamma d^2}{D}$$

$$10 \text{ s}^{-1} < \gamma < 10^3 \text{ s}^{-1}$$

$$33 < Pe < 340$$

$$D = \frac{kT}{6\pi\mu R}$$

Virus

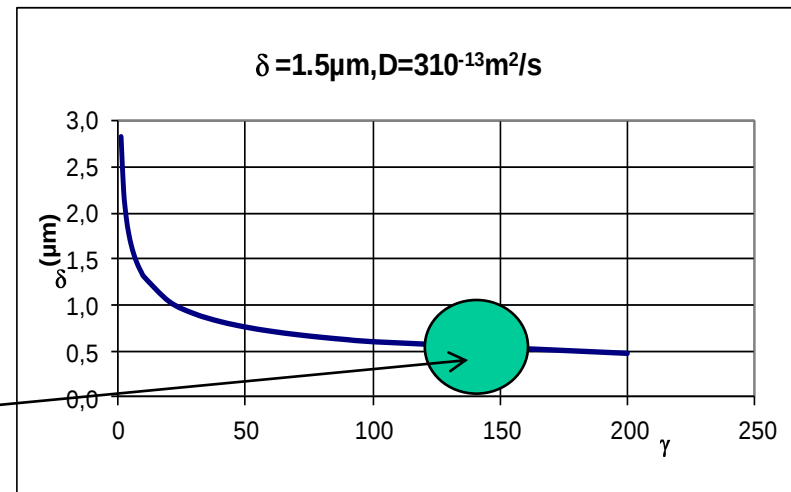
$$D \approx 1.410^{-11} \text{ m}^2 / \text{s}$$

$$0.0007 < Pe < 0.07$$

Diffusion boundary layer

$$\delta^* = \delta \sqrt{\frac{\gamma}{\nu}} = 3.68 \sqrt{\frac{D}{\nu}} Pe^{\frac{1}{6}}$$

Bacteria



Transport equation

$$\vec{J} = C\vec{V} - D\vec{\nabla}C$$

$$\frac{\partial C}{\partial t} + \text{div} \vec{J} = 0 \Rightarrow \frac{\partial C}{\partial t} + \vec{V} \cdot \vec{\nabla} C = D \Delta C$$

$$\frac{\partial C}{\partial t} + \vec{V} \cdot \vec{\nabla} C = D \Delta C \begin{cases} C = 0 & \text{for } y = 0 \text{ (activewall)} \\ C = C_0 & \text{for } y \rightarrow \infty \text{ (bulk)} \\ \left. \frac{dC}{dy} \right|_{y=0} = 0 & \text{for } y = 0 \text{ (Inert wall)} \end{cases}$$

Transport equation

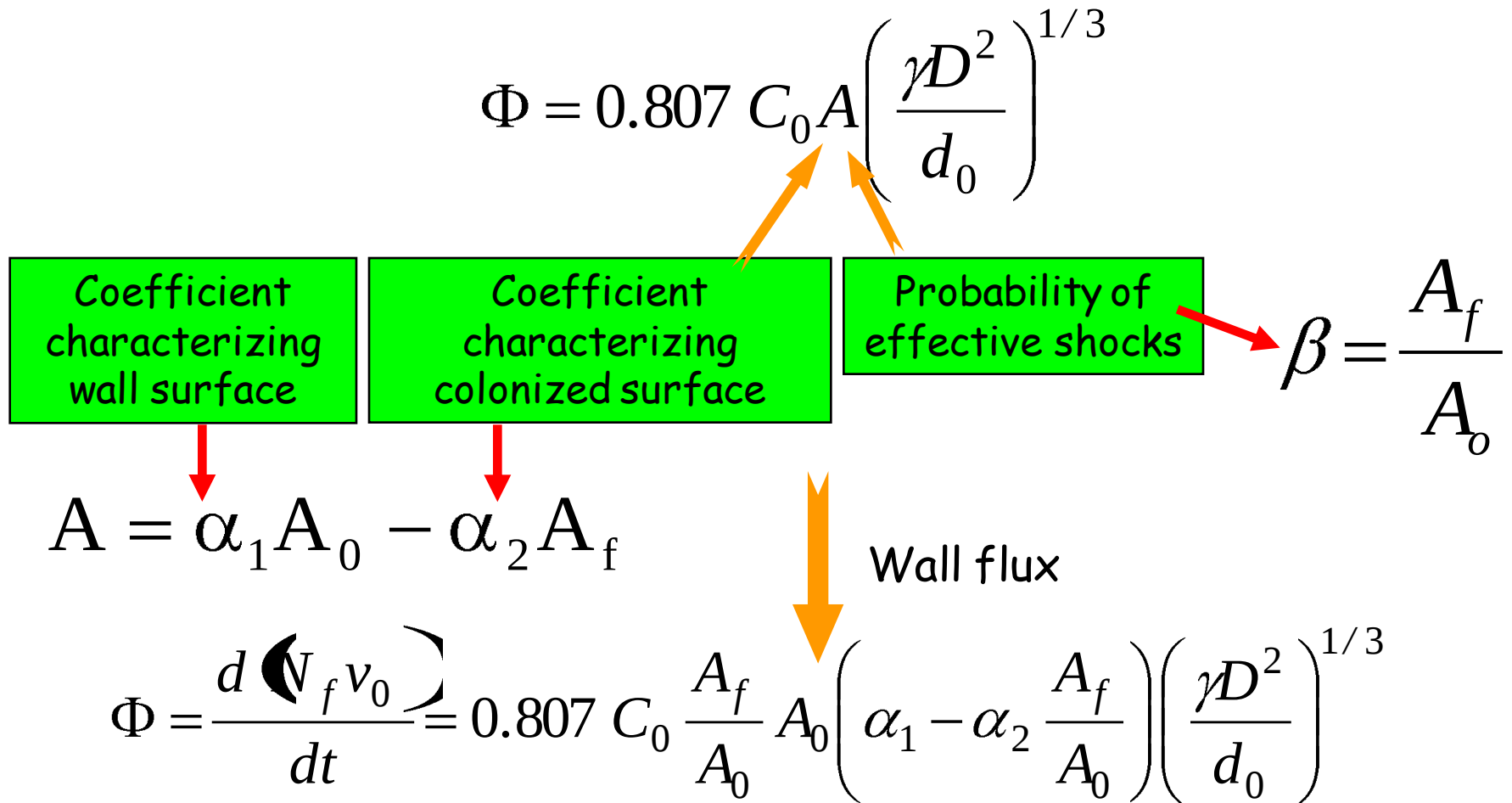
In permanent regime

$$y\gamma \frac{\partial C}{\partial x} - \frac{y^2}{2} \frac{d\gamma}{dx} \frac{dC}{dy} = D \frac{d^2 C}{dy^2}$$

$$\frac{C}{C_0} = \frac{1}{\Gamma(4/3)} \int_0^\eta \exp(-\xi^3) d\xi \quad \text{avec} \quad \eta = y\sqrt{\gamma} \left[9D \int_0^x \sqrt{\gamma} dx \right]^{1/3}$$

$$\Phi = \int D \frac{dC}{dy} \Big|_{y=0} dA = \frac{3/2}{\Gamma(4/3)} \frac{C_0 D L}{9^{1/3}} \left[\frac{\gamma d_0^2}{D} \right]^{1/3} = 0.807 C_0 A_0 \left(\frac{\gamma D^2}{d_0} \right)^{1/3}$$

Transport equation



Transport equation

The number of bacteria/unit surface (N^*) deposited is related to an adimensional time t^*

$$t^* = \left(\frac{\gamma D^2}{d_0} \right)^{1/3} t$$

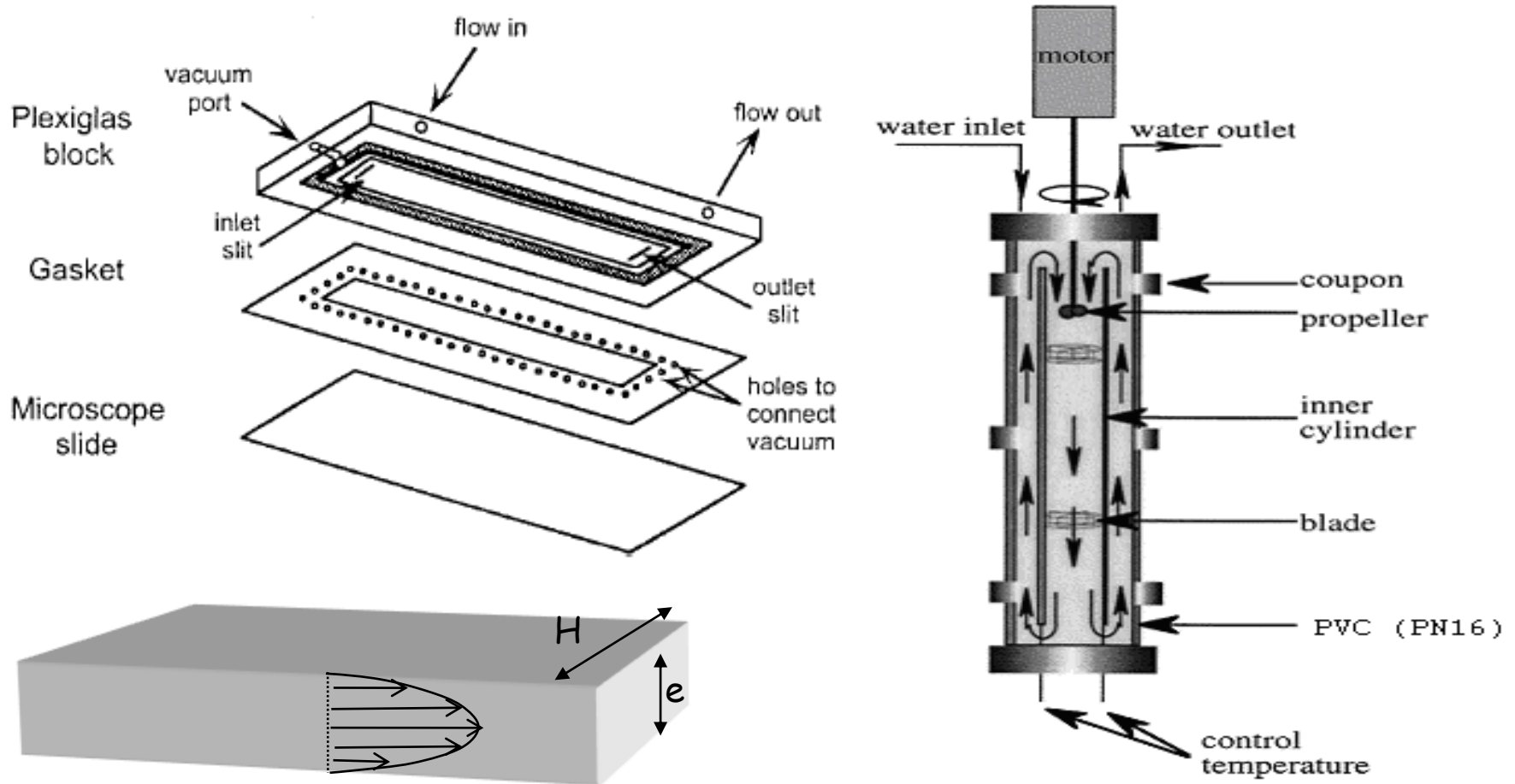
$$N^* = \frac{\frac{\alpha_2 s_0}{\alpha_1} + s \exp \left[-0.807 \frac{\alpha_1 s_0 C_o}{v_0} t^* \right]}{1}$$

Characterize the saturation

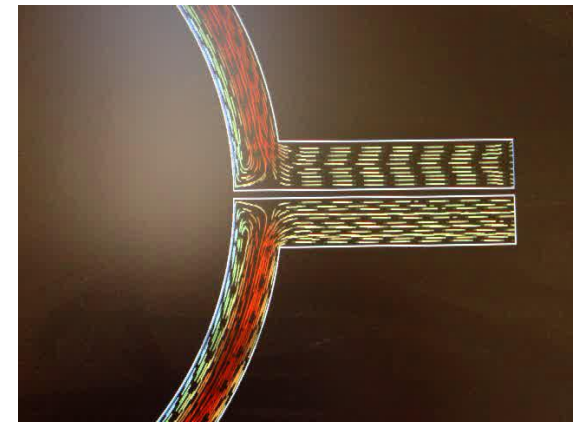
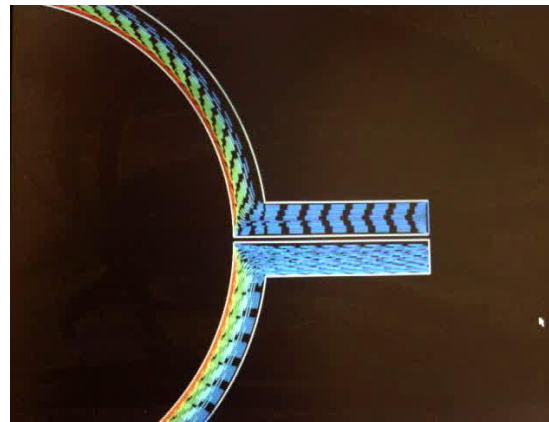
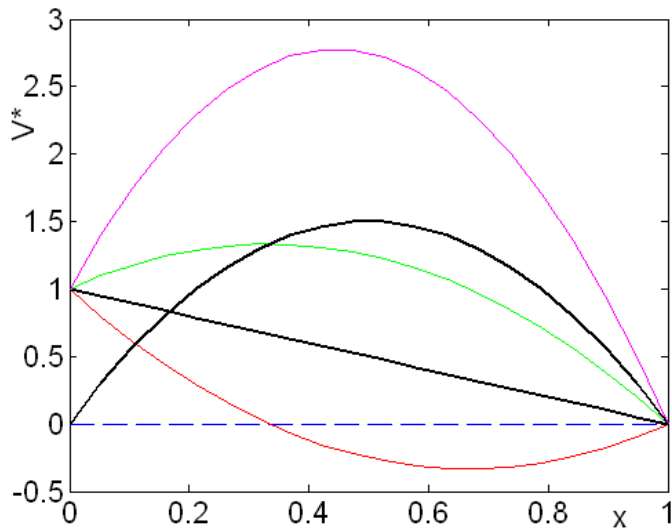
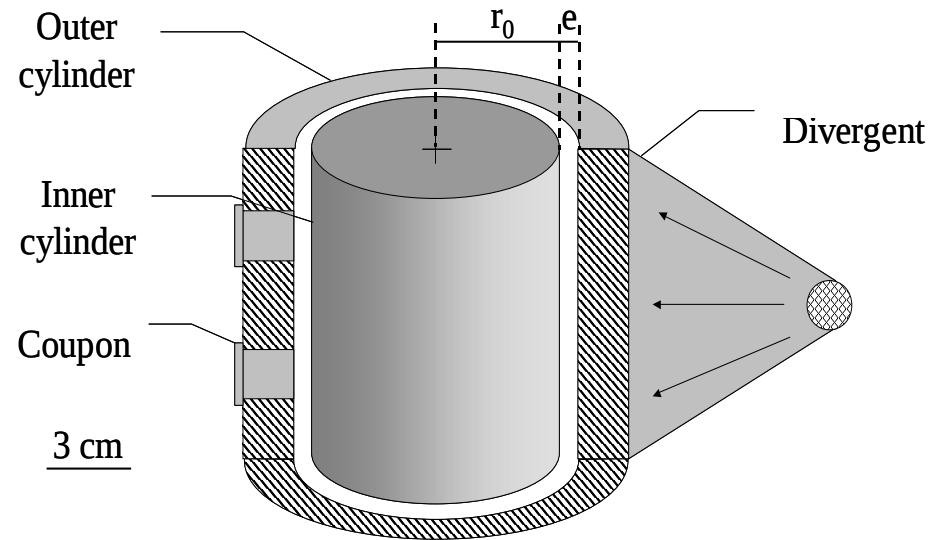
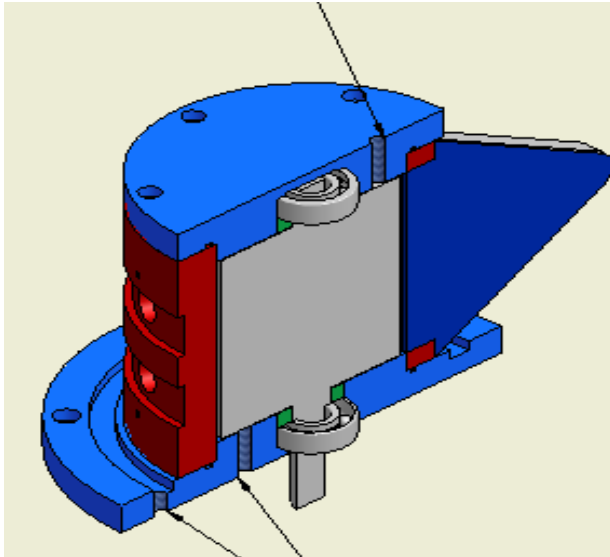
Characterize the initial state of the wall

Characterize the accumulation kinetic

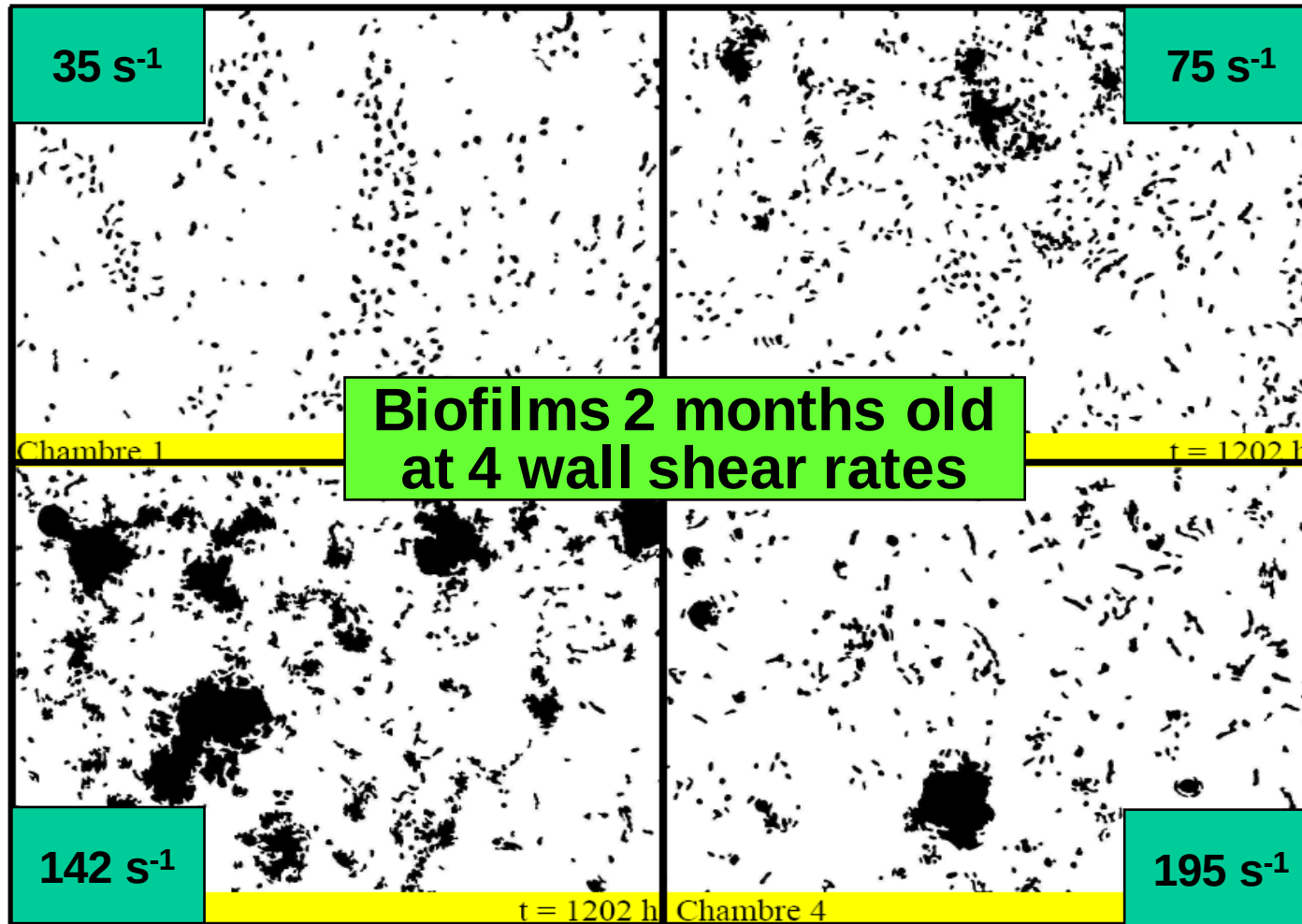
Test sections



Test sections

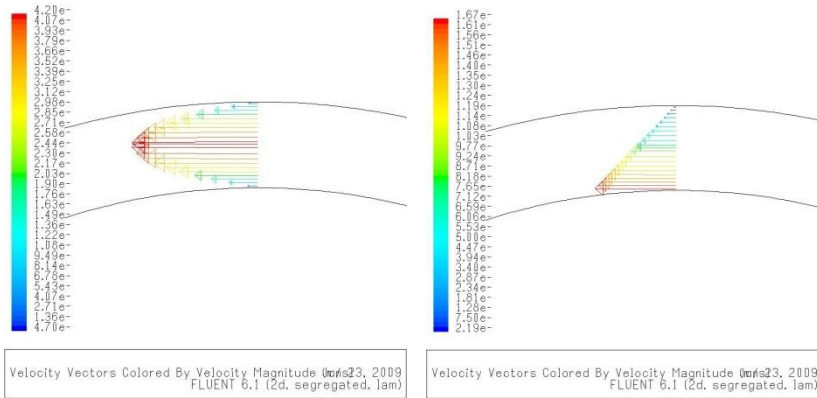


Test sections

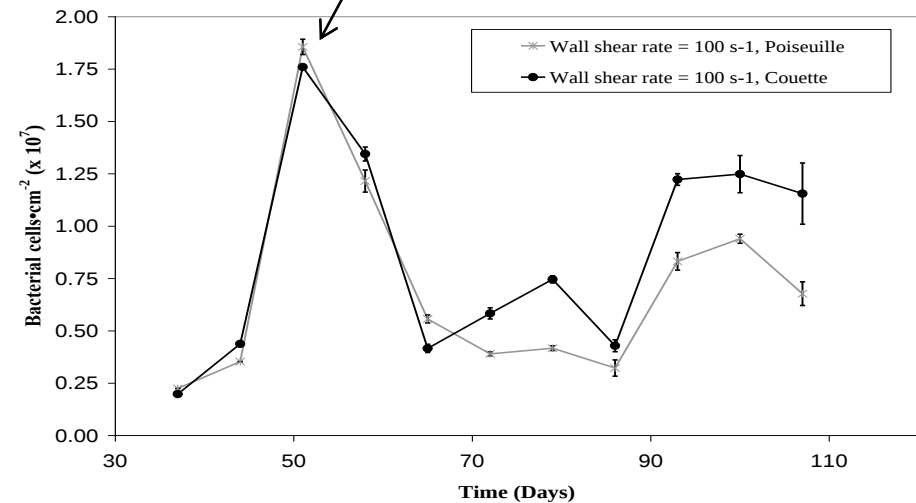
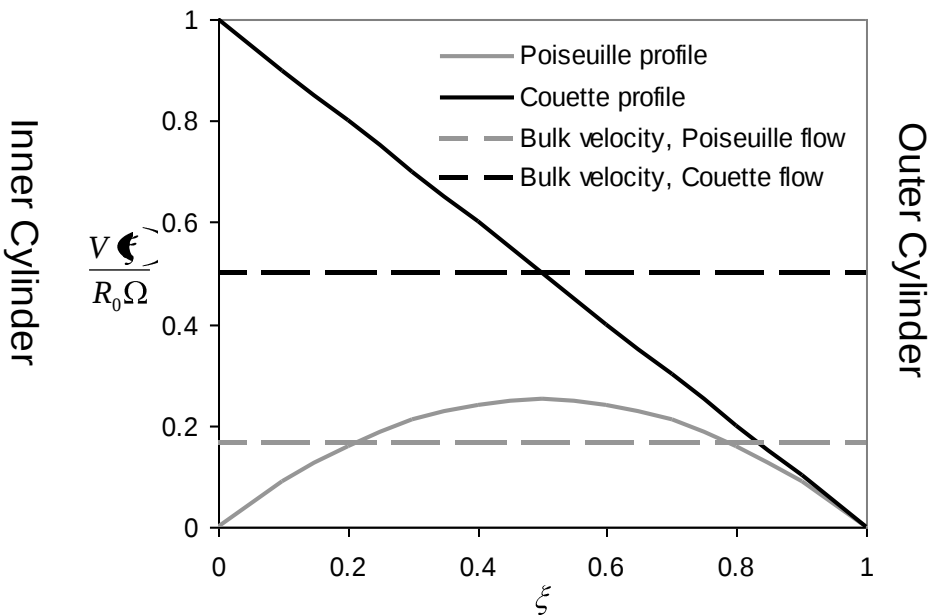


Increasing the wall shear rate led to a space organization of bacteria deposition

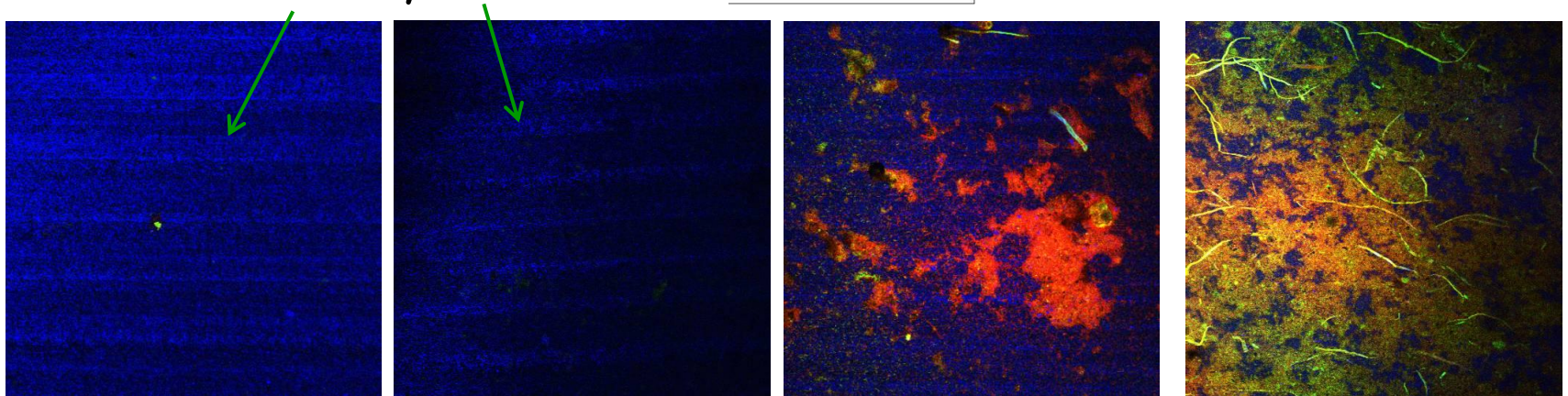
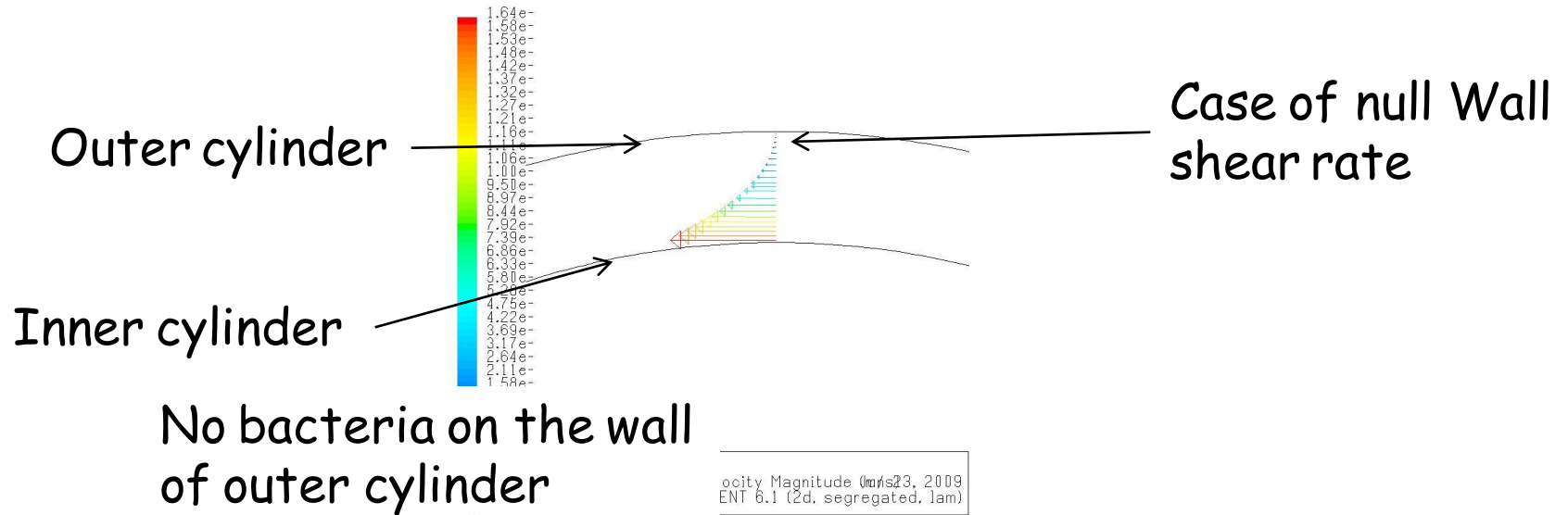
Test sections



Grazing by Amoeba



Test sections



Conclusions on wall flux

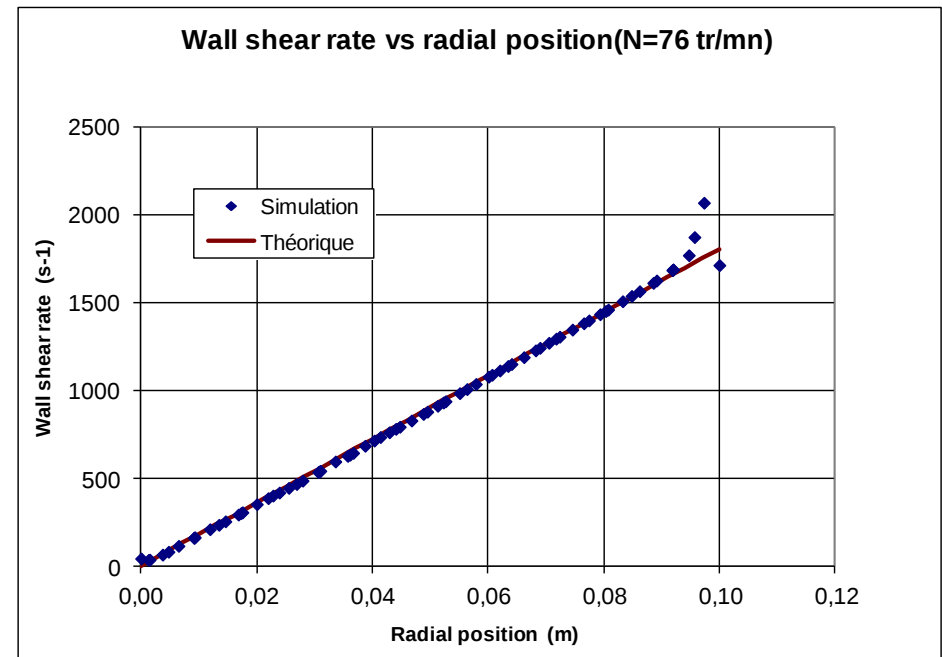
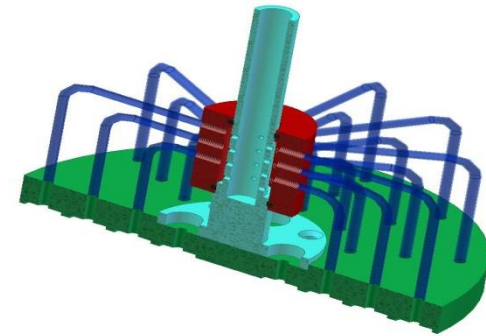
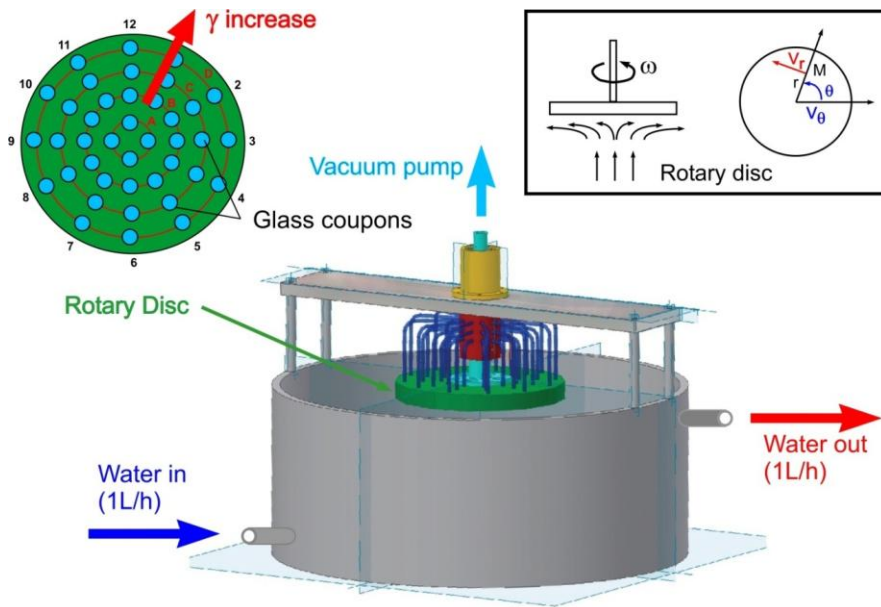
Bacteria in drinking water : migration to the wall -> biofilm ?

No wall shear rate -> no bacteria on the wall

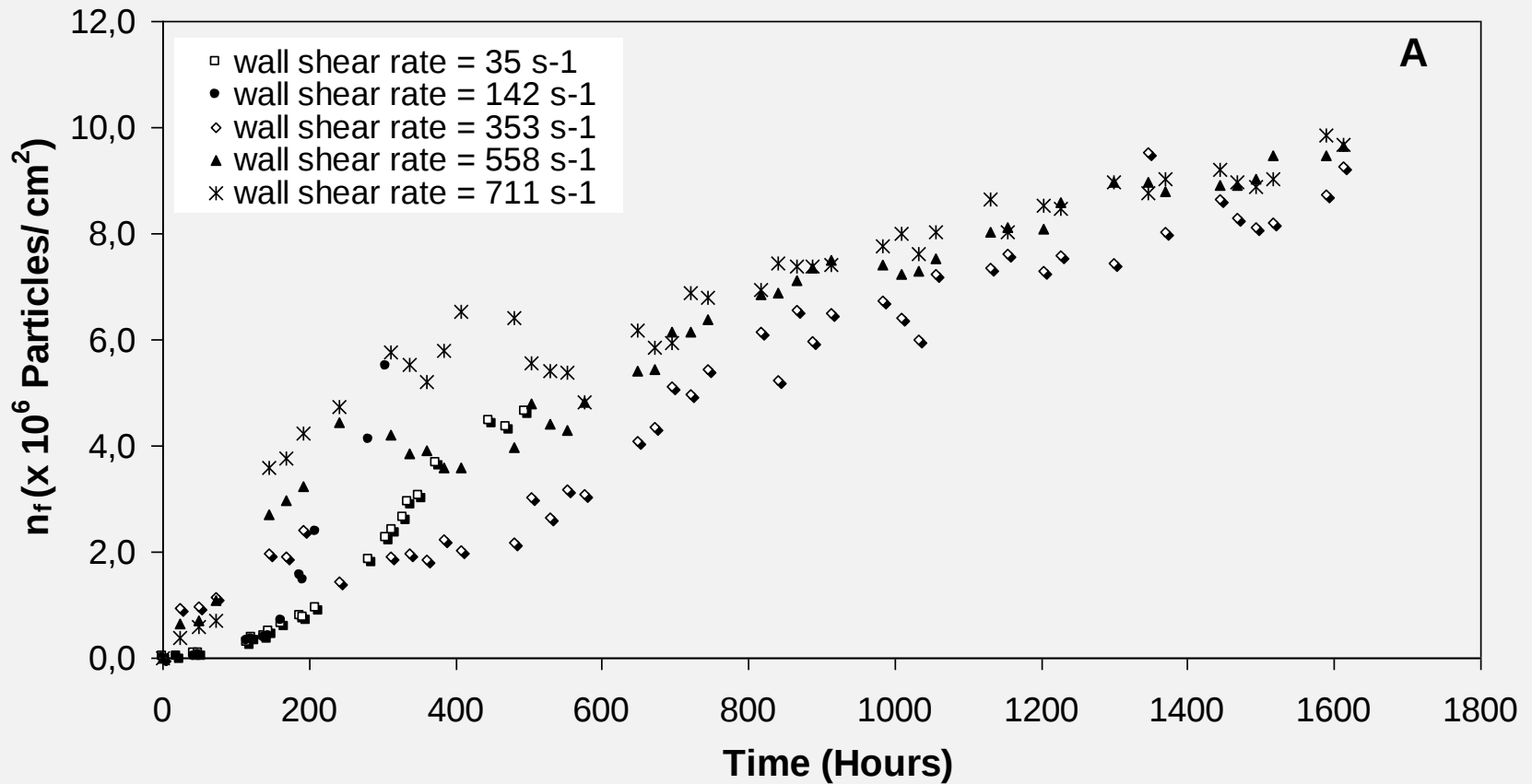
Even if the Peclet number is high -> Convection –diffusion works

$$N^* = \frac{1}{\frac{\alpha_2 s_0}{\alpha_1} + \exp\left[-0.807 \frac{\alpha_1 s_0 C_o}{v_0} t^*\right]}$$

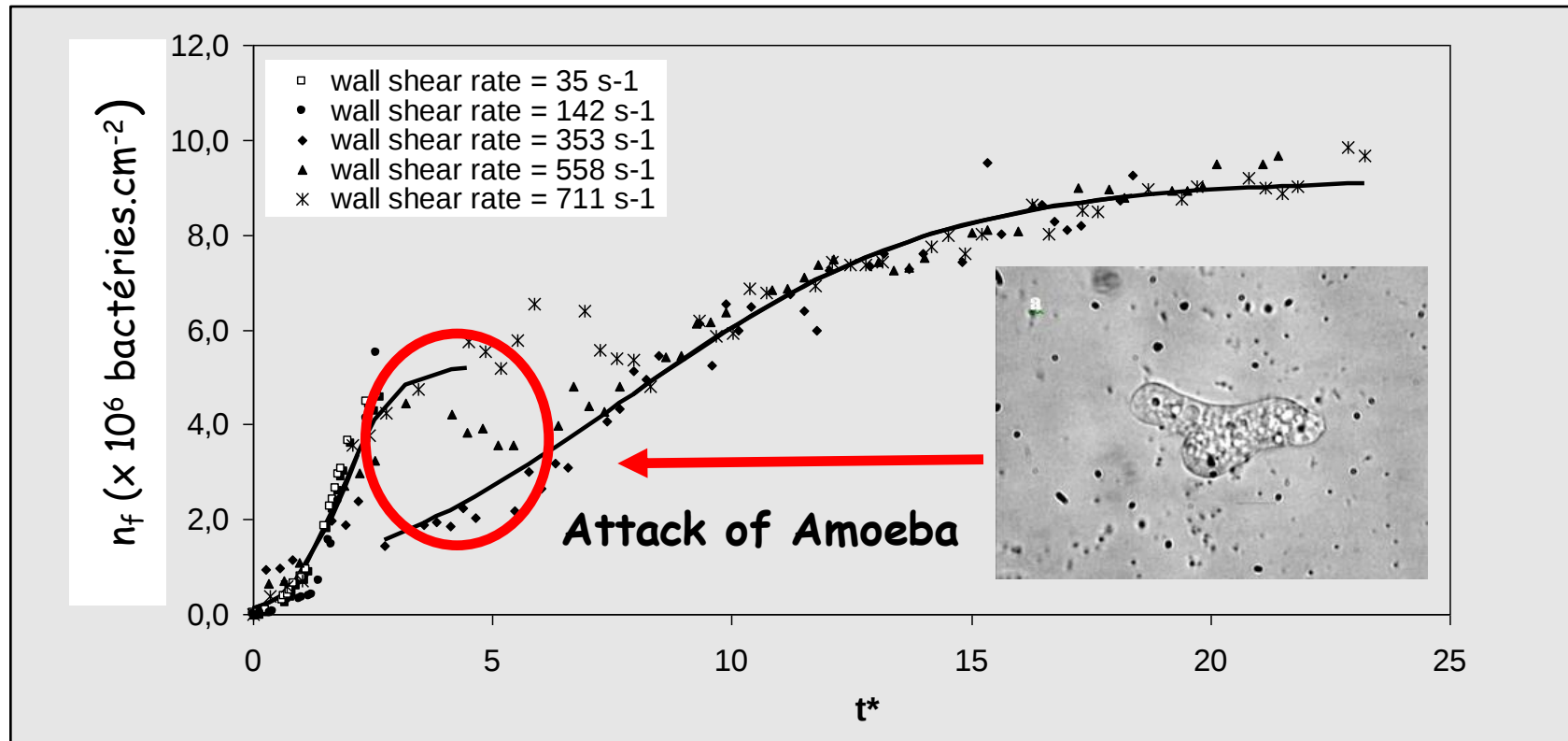
Test sections (Rotating disc)



Results

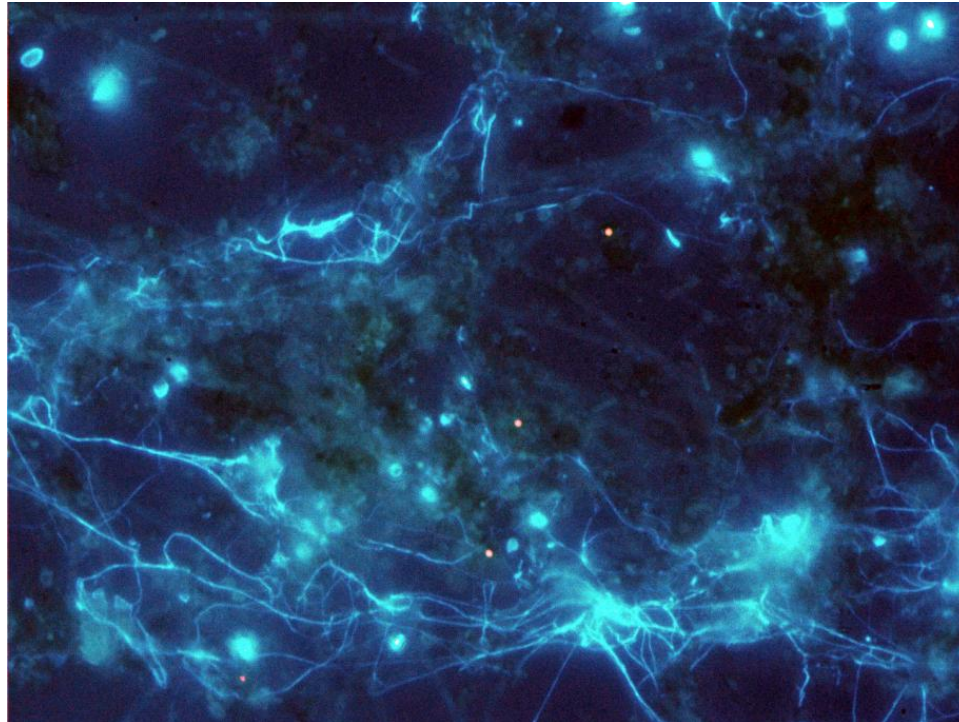


Results



	Phase 1	Phase 2
alpha 1	1	0,3
alpha 2	30	5
S_0	$8 \cdot 10^{-6}$	$1 \cdot 10^{-6}$

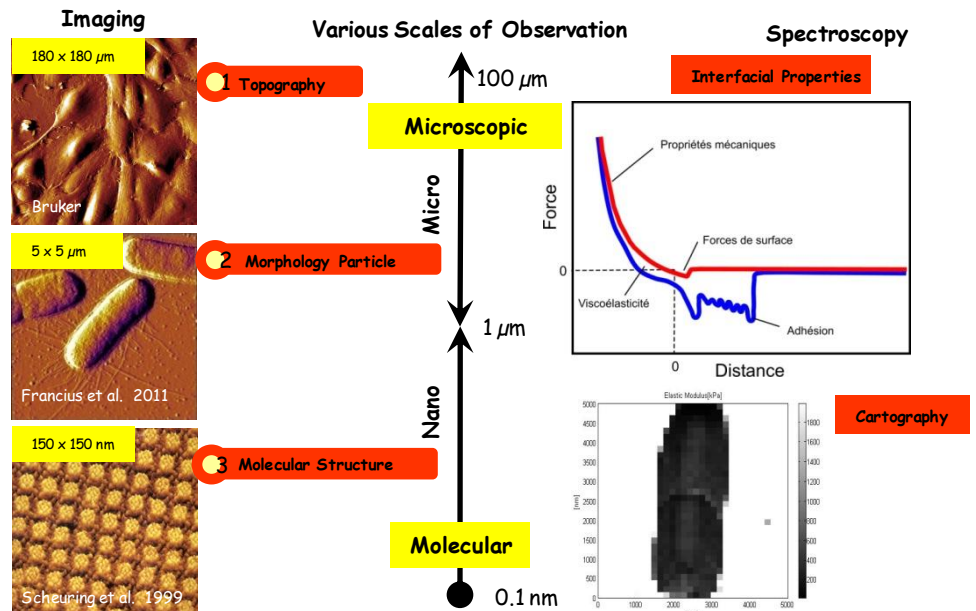
Bacteria adhesion at the wall (cohesion of biofilm)



Measurement technique

AFM $\Rightarrow$ an application of the scanning tunneling microscope (STM) :

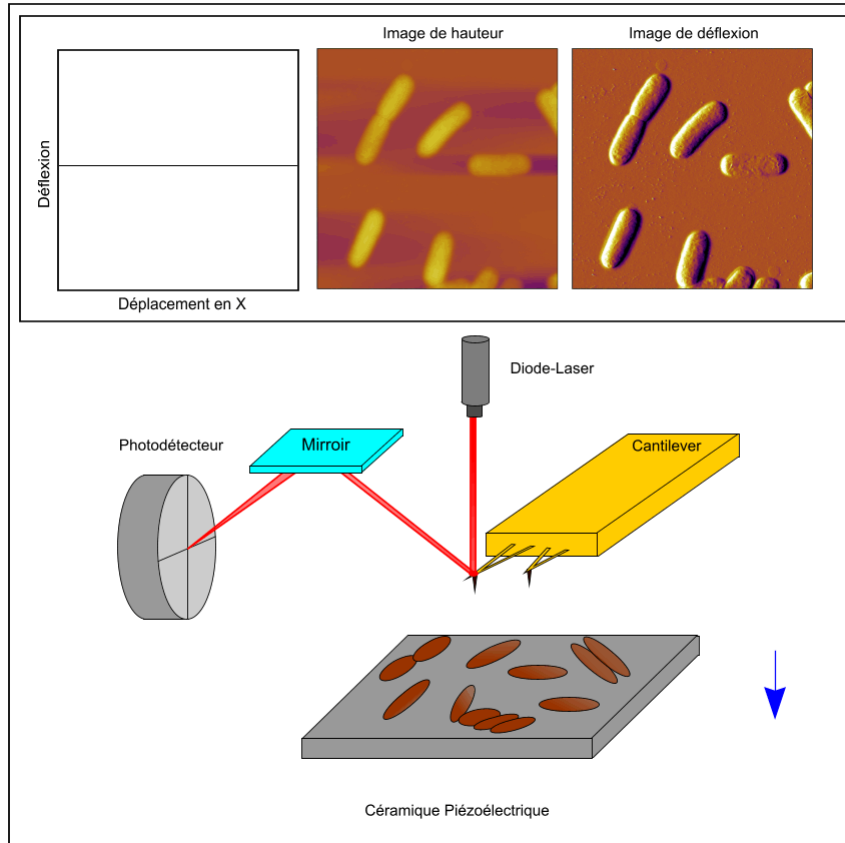
➤ Imaging samples surface in various environments



In the last decade $\Rightarrow$ experiencing boom of AFM in nanosciences and life sciences :

AFM technique

Panorama of measurable forces in AFM



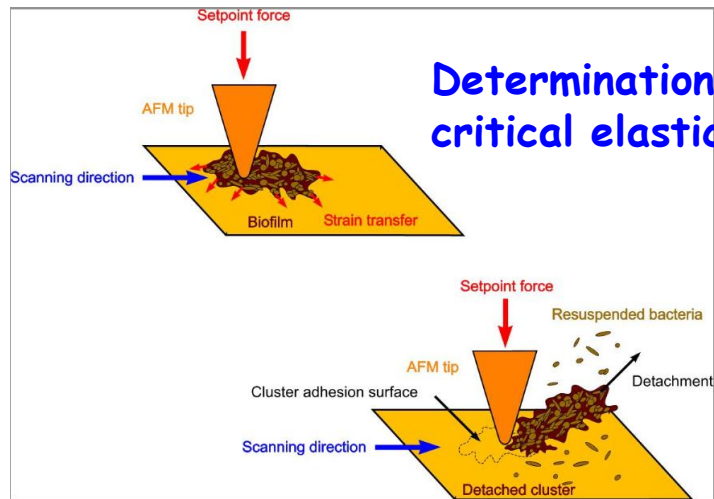
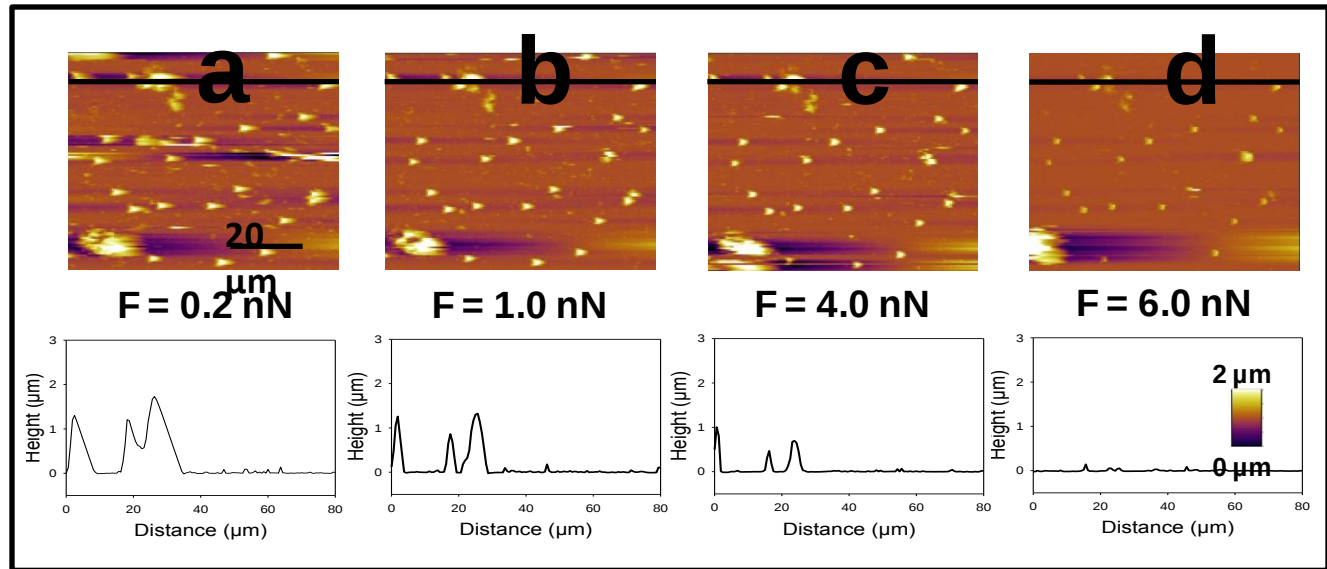
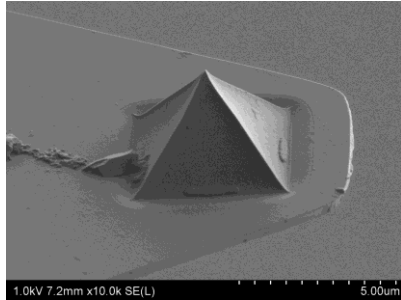
Approach		Retraction			
a van der Waals	$F(D) = \frac{AR}{12D^2}$	e Adhesion	$F = -3\pi R\gamma$		
b Electrostatic	$F(D) = \frac{4\pi R\lambda\sigma_p\sigma_s}{\epsilon} e^{-D/\lambda}$	f Capillary force	$F = 4\pi R\gamma_l \cos\theta$		
c Brush	$F(D) \approx \frac{50LkT}{s^3} e^{-2\pi DL}$ $0.2 \leq D/2L \leq 0.9$	g Polymer extension	$F(x) = \frac{kT}{a} L^* \left(\frac{x}{Na} \right)$		
d Elastic	$F(\delta) = \frac{4EVR}{3(1-\nu^2)} \delta^{3/2}$	h Binding	$F = \frac{U - kT \ln(\tau/\tau_0)}{\Lambda}$		
Definitions					
A	Hamaker constant	T	Absolute temperature	Λ	Characteristic length of bond
a	Monomer length	U	Bond energy	λ	Debye length of the medium
D	Probe-sample separation distance	x	Elongation of polymer	θ	Angle related to the geometry of the tip-sample contact
E	Elastic modulus	δ	Indentation depth	σ_R	Surface-charge density of sphere
k	Boltzmann's constant	ϵ	Dielectric of the medium	σ_S	Surface-charge density of sample
L	Brush thickness in a good solvent	γ	Surface energy between tip and sample	τ	Period over which the bond will rupture
L*	Inverse Langevin function	γ_L	Surface energy of the liquid	τ_0	Reciprocal of the natural bond frequency
N	Number of units in polymer	s	Mean distance between polymers		
R	Radius of probe sphere	v	Poisson ratio		

Atomic force spectroscopy:

- Combination of imaging and force measurements
- Mapping reconstruction with spatially resolved physical parameter value for each pixel
- Which physico-chemical parameters can be extracted from the AFM force-curves?

AFM : Cohesion of drinking water biofilm clusters

Scanning surface
by AFM ($F \uparrow$)

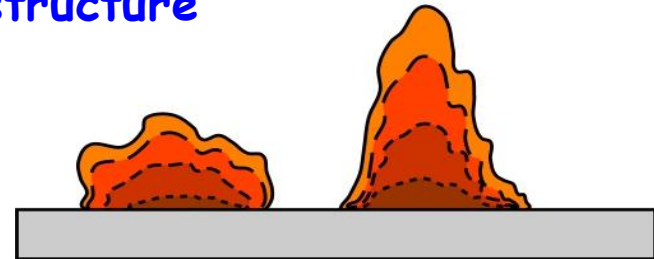


Determination of the
critical elastic modulus

Young modulus

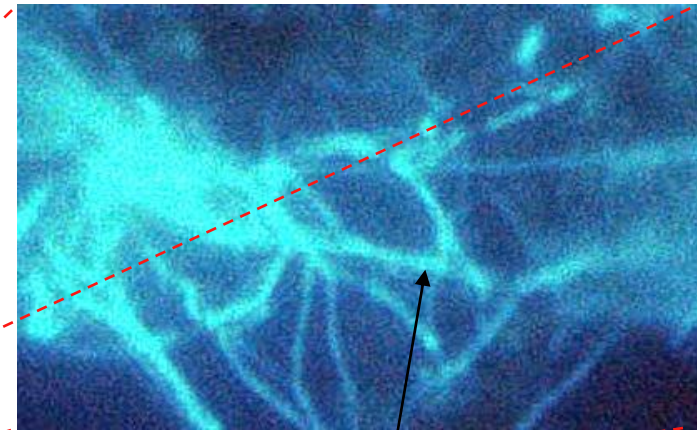
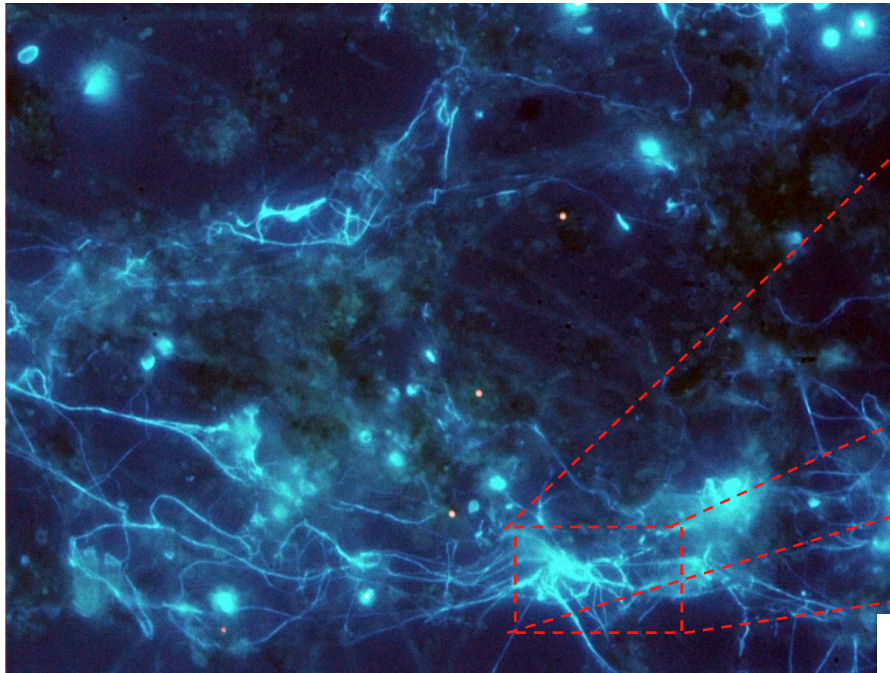
0 MPa  10 MPa

Stratification of the cluster
structure

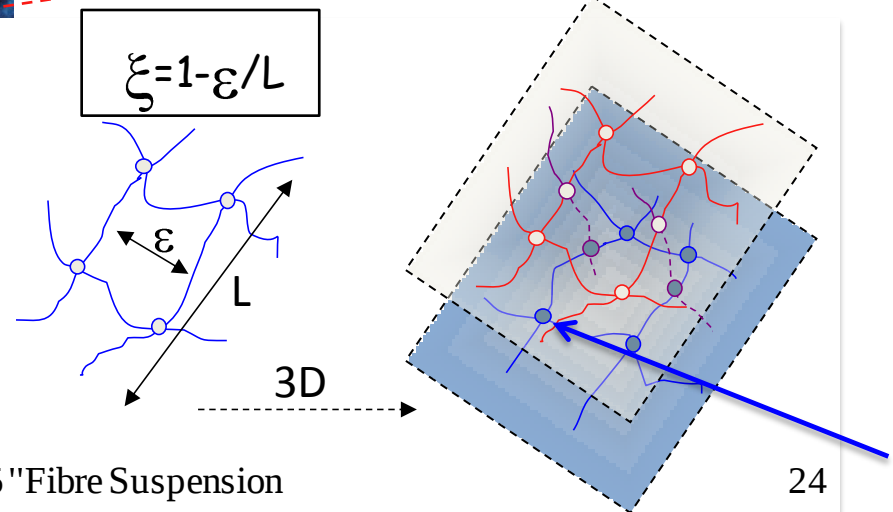


Water drinking biofilm

The rate of entanglement ξ is a factor of cohesion linked to the volume and elasticity of the clusters



G. De Gennes
 ε : coherence length



AFM : Cohesion of clusters in biofilm drinking water

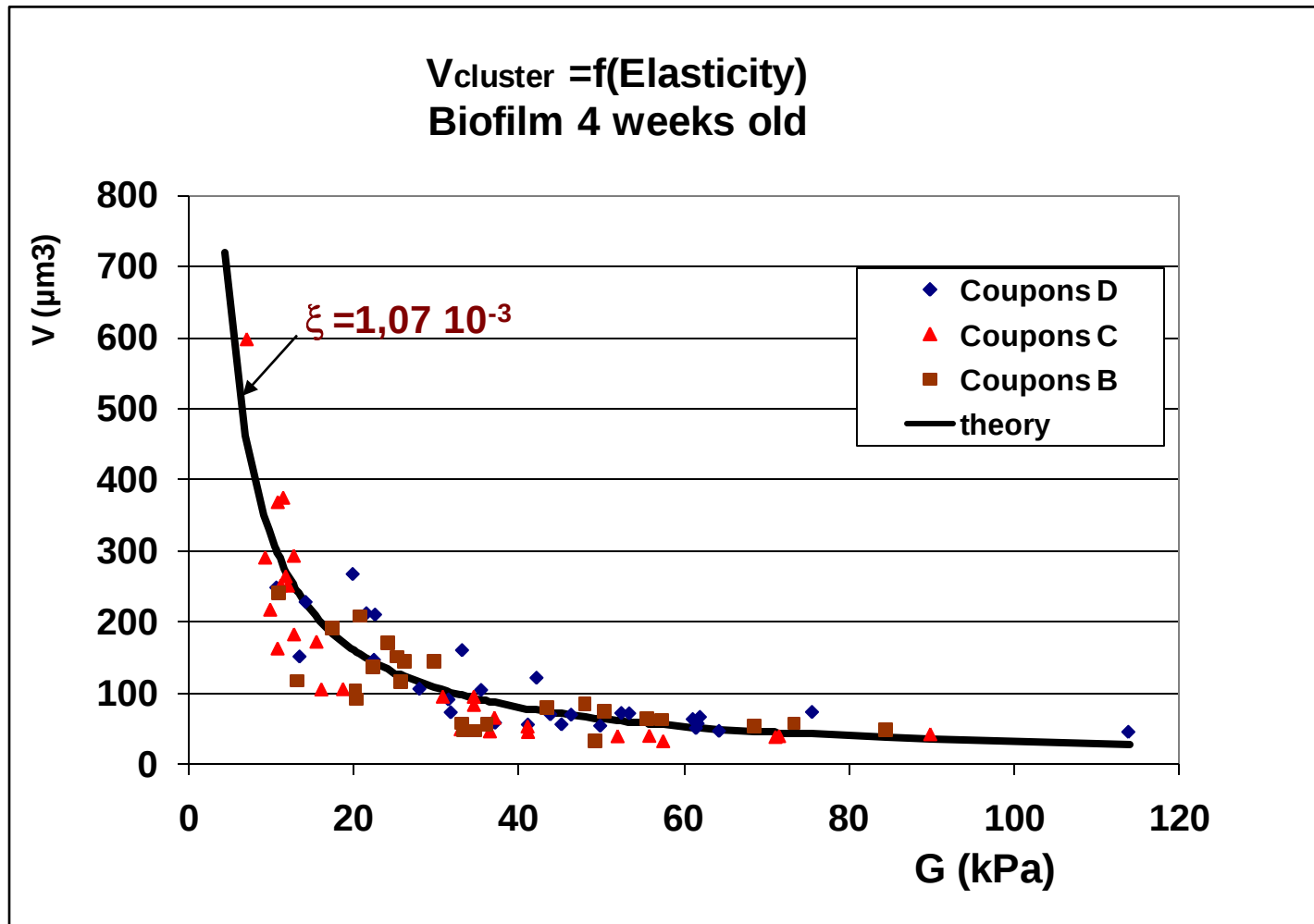
$$W_{elas} \approx G L^3 \approx G \left(\xi d \right)^3 \approx G \xi^3 V_{cluster} \quad avec \quad \begin{cases} G : \text{Shear modulus elasticity} \\ \xi : \text{rate of entanglement in cluster} \\ V_{cluster} : \text{volume of cluster} \end{cases}$$

$$W_{elas} = k_B T \quad avec \quad \begin{cases} k_B = 1.38 \cdot 10^{-23} \text{ m}^2 \text{ kg/s}^2 / ^\circ \text{K} \text{ Boltzmann constante} \\ T : \text{absolute temperature} \end{cases}$$

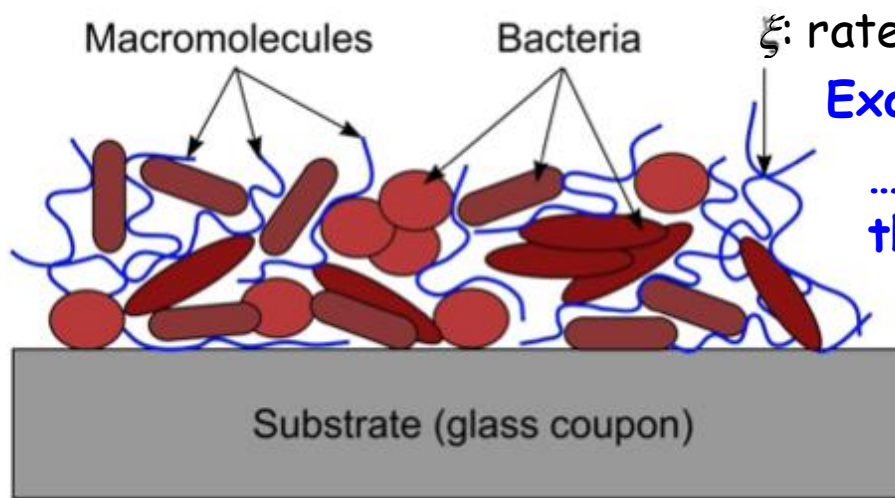
At the limit of cohesion, cluster we can put:

$$G \xi^3 V_{cluster} = k_B T \Rightarrow \boxed{V_{cluster} = \frac{k_B T}{\xi^3 G}} \quad \begin{aligned} &k_B T = 4 \cdot 10^{-21} \text{ m}^2 \text{ Kg/s}^2 / \text{K} \\ &V_{cluster} \text{ and } G \text{ given by AFM} \end{aligned}$$

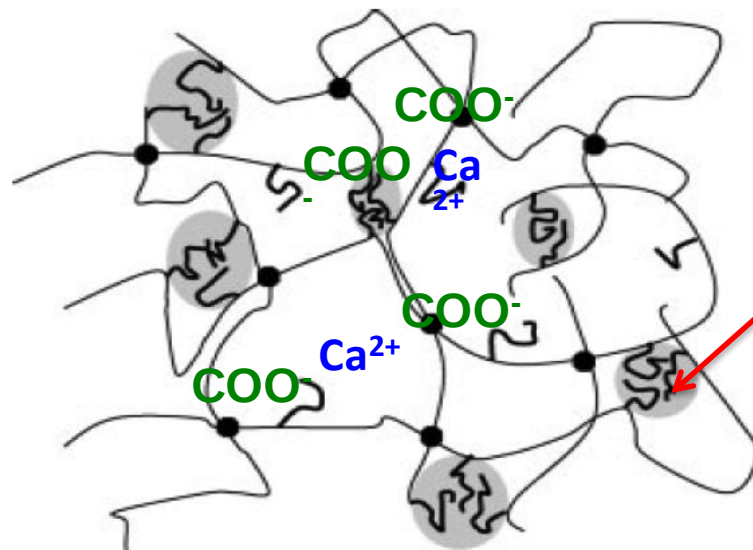
AFM : Cohesion of clusters in biofilm drinking water



Cohesion of clusters in biofilm drinking water

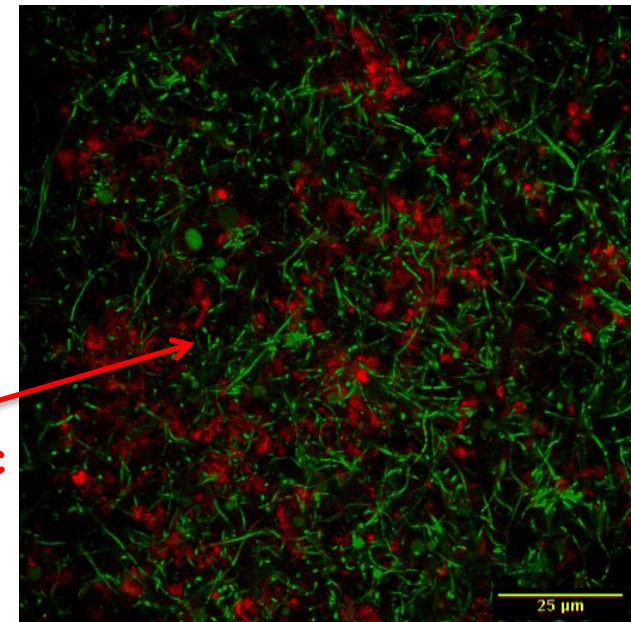


Exopolymers in drinking water biofilms ...
... govern the elastic deformation of the clusters



(Miquelard-Garnier *et al.*, 2007)

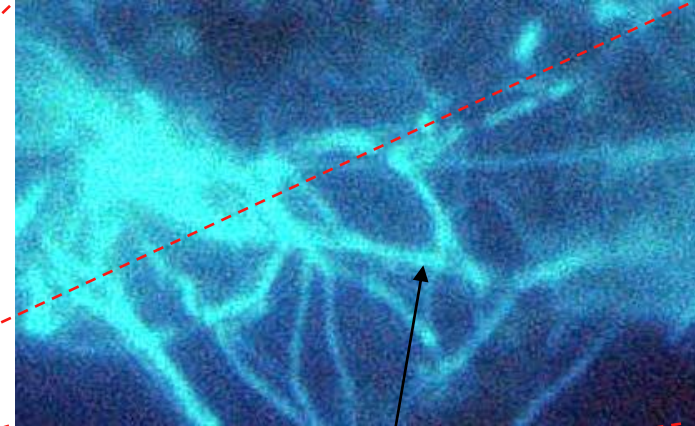
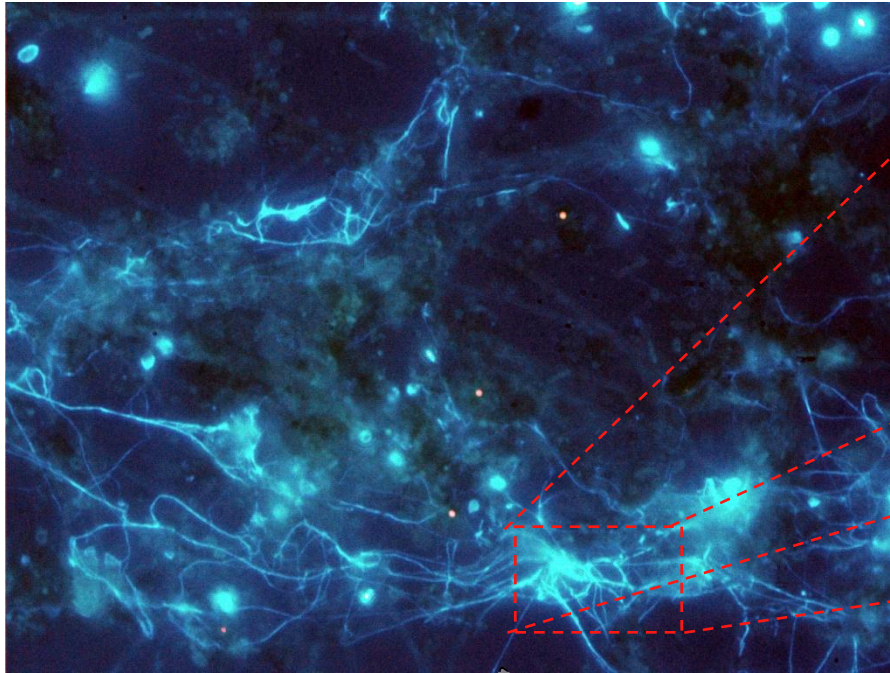
Hydrophobic domains



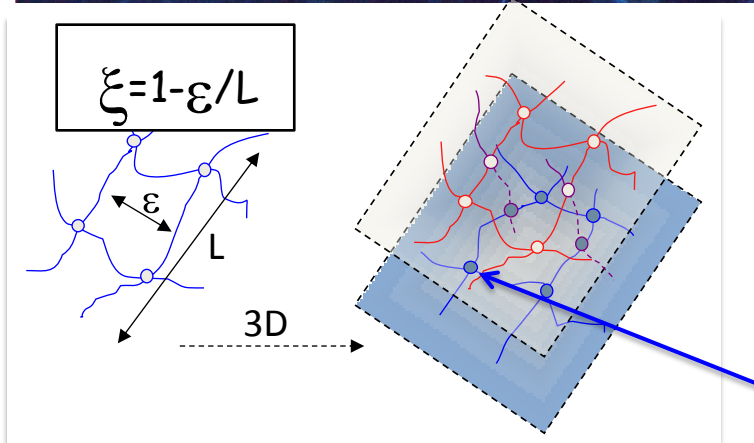
Aldeek *et al.*, AEM, 2013
(Biofilm of *Shewanella oneidensis*)

Number of entanglement points $10^9/\text{mm}^3$

The rate of entanglement ξ is a factor of cohesion linked to the volume and elasticity of the clusters



$$V_{cluster} = \frac{k_B T}{\chi^3 G}$$



Drinking water biofilms behave like a viscoelastic solid

$$\xi = 10^{-4} \text{ to } 10^{-3}$$

Number of entanglement points 10^8 to $10^9/\text{mm}^3$

Cleaning the surface colonized by biofilm

hydrodynamic shear stress vs cluster volume

Mean shear stress in volume of cluster V: $\sigma_{ij} = \frac{1}{V} \sum_1^n \left(\int_{A_0 - A_1} \sigma_{ik} x_j n_k dA + \int_{A_1} \sigma_{ik}^b x_j n_k dA \right)$

$$\sigma_{ij} = \sigma_{ij}^f C(1 - \xi) + \sigma_{ij}^b C \xi$$

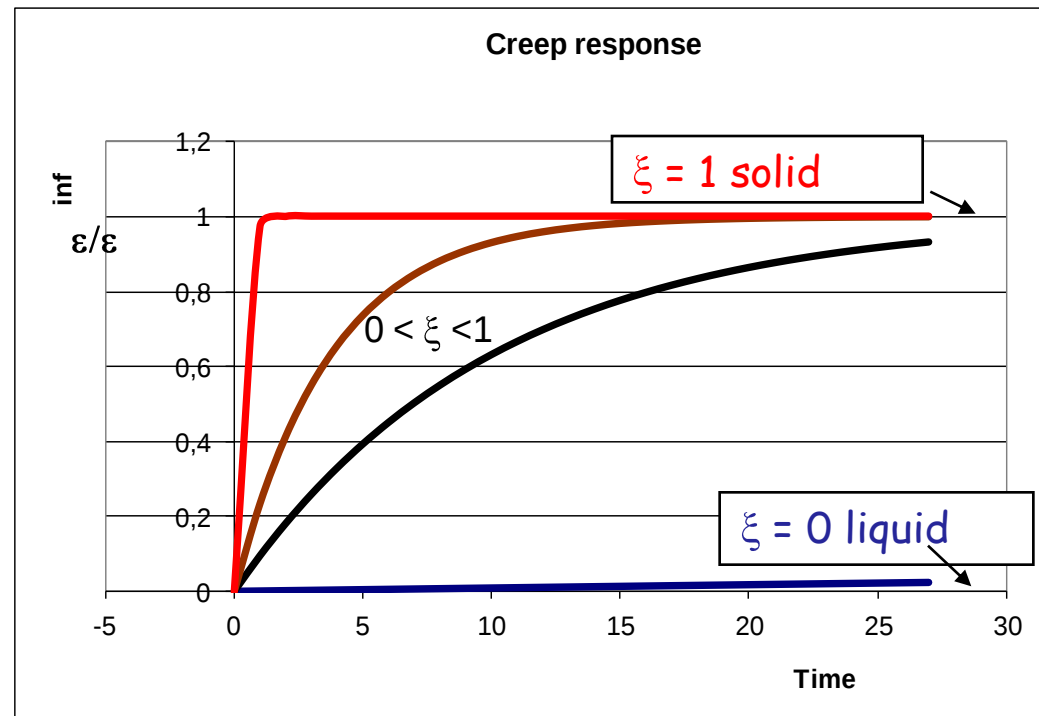
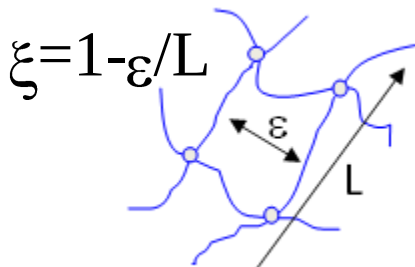
$$\sigma_{ij}^f = \mu \frac{d\alpha}{dt} = \mu \gamma \quad \text{et} \quad \sigma_{ij}^b = G \alpha$$

$$\tau = \mu C(1 - \xi) \frac{d\alpha}{dt} + G C \xi \alpha$$

Creep function

$$f(t) = \frac{1}{G \xi} \left(1 - e^{-\frac{t}{\theta}} \right)$$

with $\theta = \frac{\mu(1 - \xi)}{G \xi}$



Cleaning the surface colonized by biofilm

hydrodynamic shear stress vs cluster volume

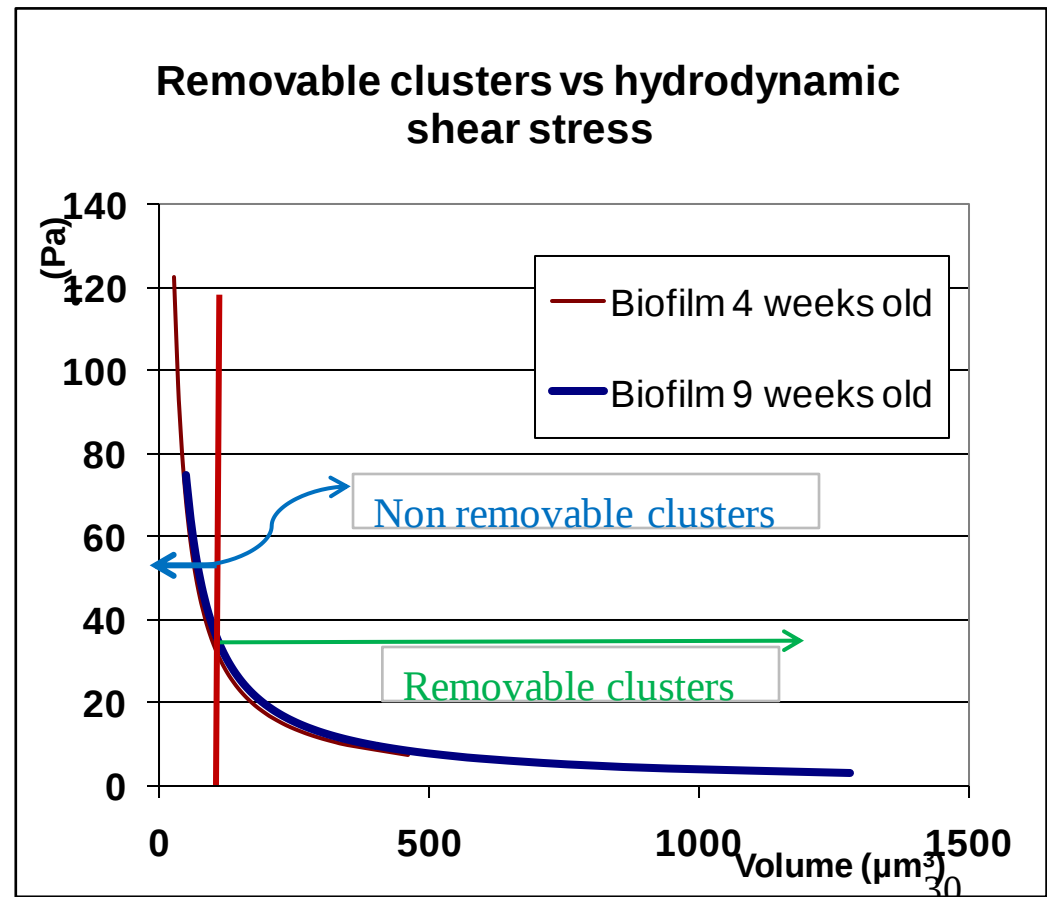
Elastic limit before removing clusters:

$$\mu \frac{d\alpha}{dt} C (1 - \xi) = \mu \gamma C (1 - \xi) = G C \xi \alpha_{\max}$$

Strain rate $\alpha_{\max} = 100\%$:

$$\tau_{hyd} = \mu \gamma = \frac{G \xi}{1 - \xi}$$

City network drinking water
distribution $\tau < 30$ Pa



Conclusions

- Biofilm formation –convective diffusion
- Strong adhesion at the wall (AFM measurements)
- Cleaning the surface colonized by biofilm with only hydrodynamic shear stress -> volume clusters $<100\mu\text{m}^3$ non removable
- Perspective : diffusion of nutrients on biofilm - population balance (growth, mortality and partial detachment)

THANK YOU FOR YOUR ATTENTION

- Tony Paris, Salaheddine Skali-Lami, Jean-Claude Block (2007) : ***Effect of Wall Shear Rate on Biofilm Deposition and Grazing in Drinking Water Flow Chambers***. Biotechnology and Bioengineering, V. 97, issue. 6: 1550_1561.
- Paris T., Skali-Lami S., and Block J.C. (2009) : ***Probing young drinking water biofilms with hard and soft particles***. *Water Research*, v. 43, issue 1: 117 – 126
- El Khatib R., Lartiges B., Mustin C., Skali-Lami S. (2011) : ***Hydrodynamic control of wastewater biofilm organization***. *La Houille Blanche*, Issue 5: 29 – 33.
- Abe Y., Polyakov P., Skali-Lami S., and Francius G. (2011) : ***Elasticity and physico-chemical properties during drinking water biofilms formation***. *Biofouling*, v. 27(7): 739 – 750.
- Abe Y., Skali-Lami S., Block J.C., and Francius G. (2012) : ***Cohesiveness and hydrodynamic properties of young drinking water biofilms***. *Water Research*, 46: 1155 – 1166.

